

**Research Article****Effect of Gonadotrophin (Diclair®) on Semen Characteristics, Testicular Morphometry and Hormonal Profile of Mature Harco Cocks**

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Article History: Received: July 12, 2016 Revised: August 16, 2016 Accepted: September 05, 2016**ABSTRACT**

Twenty sexually matured (24 weeks old) healthy Harco cocks were used to determine the effect of Gonadotrophin (Diclair®) on semen characteristics, testicular morphometry and hormonal profile. The cocks were divided into 4 treatment groups of 5 cocks per group identified as T₁ (control) administered with 1ml physiological saline, T₂, administered with 13.50i.u Diclair®, T₃ 27.00i.u and T₄, administered with 40.50i.u Diclair®, with one cock per replicate in a Completely Randomized Design (CRD). The injections were divided into three doses each and administered intramuscularly in the thigh for three consecutive days. Semen was collected one week after Diclair® administration, twice a week for 4 weeks by the abdominal massage and manipulation of the cloaca method. Five cocks were randomly selected from each treatment group and bled one week after Diclair® injections to collect blood for hormonal profile evaluation. The results showed that there were significant differences ($P < 0.05$) among the treatment groups in all the semen parameters; semen volume, semen pH, individual motility, sperm concentration, proportions of dead sperm cells and live sperm cells. The results further showed that there were significant differences ($P < 0.05$) among the treatment groups in testicular, epididymal and vas deferens weights. Similarly, the results showed significant differences ($P < 0.05$) among the treatment groups in follicle stimulating hormone (FSH), luteinizing hormone (LH) and testosterone levels. The results of this study suggest that Diclair® treatment enhanced semen quality, testicular development and was not detrimental to the hormonal profile of the cocks.

Key words: Diclair® semen quality, Testicular morphometry, Hormone, Cocks**INTRODUCTION**

Harco chicken was developed in New England in the middle of the 19th century and was first exhibited as a breed in 1869. The Harco chicken is a dual-purpose, cold-hardy bird and therefore makes a great breed for the small farm or backyard flock owner. Harco cocks are large, long-lived chicken weighing between 2.5kg-2.8kg and are bred principally for meat. They possess a long broad back, a moderately deep, full breast, yellow skin and legs. The birds are dark barred in colour which means that the bars of dark colour are wider than the white colour.

In order to carry out any sustainable improvement in livestock, there should be methods of ensuring the repeatability and multiplication of desired traits in subsequent generations. Reproduction is a process by which an organism gives rise to a new member of its species. It is a vital factor in determining the efficiency of animal production and its performance is closely related to profitability in poultry enterprise (Iheukwumere *et al.* (2008).

Sperm producing potentials are evaluated by aspects of semen output: volume, motility of sperm cells, morphology of spermatozoa, proportion of live sperm cells and concentration in ejaculate. No single parameter has been proved to be an accurate predictor of the quality of individual ejaculates (Iheukwumere *et al.*, 2001). Sperm formation involves the use of follicle stimulating hormone (FSH) and luteinizing hormone (LH) (Iheukwumere *et al.*, 2004). Most of these preparations of FSH and LH are very expensive perhaps because of the brand names, some of them require cold chain storage and often deteriorate because of inadequate storage and handling (Herbert *et al.*, 2000).

Diclair®, also known as Humegon or Mentrophin and with similar constituents as plusset® is a gonadotrophin preparation lyophilized in vials containing a mixture of follicle stimulating hormone and luteinizing hormone in a ratio 1:1 (Dixon and Hopkins, 1996). Follicle stimulating hormone and LH in Diclair® play vital role in the initiation of spermatogenesis. The hormone preparation is

cheap, readily available and does not require cold chain storage (Iheukwumere, 2005). It has not been determined if the administration of the hormone preparation for spermatogenesis and semen production would induce any side effects on the testicular morphometry and hormonal profile of the cocks. This study was therefore carried out to determine the effect of Diclair® administration on the semen quality, testicular morphometry and hormonal profile of mature Harco cocks.

MATERIALS AND METHODS

Experimental birds and their management

Twenty clinically sound and sexually matured (24 weeks old) Harco cocks purchased from Elgibbor farms in Isuikwuato Local Government Area, Abia State Nigeria, were used for this study. The birds were dewormed and vaccinated soon after purchase. A two-week pre-experimental period was allowed to enable the animals acclimatize. The birds were housed and raised on a deep-litter system. They were fed commercial Grower mash containing 20% CP and 2000 Kcal ME/kg diet twice daily (in the morning and evening). Water was provided *ad libitum*.

Experimental design

The twenty sexually matured (24 weeks old) Harco cocks were divided into 4 treatment groups, identified as T₁, T₂, T₃, and T₄. Each treatment group consisted of 5 cocks with one cock per replicate in a Completely Randomized Design (CRD), with four levels of Diclair® as treatment. The levels of Diclair® were 0.00ml, 0.09ml, 0.18ml, and 0.27ml represented as T₁, T₂, T₃ and T₄ respectively. T₁ (Treatment 1) which contained no Diclair® served as the control. Diclair® treatment was by intramuscular injection. The injections were administered as follows:

Dicclair® was supplied in 3 vials each containing FSH 75i.u and LH 75i.u. The content of the vial was dissolved in 1ml of physiological saline solution provided immediately prior to use, resulting in a solution of DFSH 75i.u plus DLH 75i.u per ml. All treatments were administered intramuscularly on the thigh of each cock using a one ml syringe with 0.01ml graduation. The doses and concentration of Dicclair® administered are shown in tables 1 and 2.

Semen collection and evaluation

The cocks were trained to ejaculate by abdominal massage and manipulation of the cloaca as described by Abu *et al.* (2006). By this method, ejaculates were collected from each cock after one week of Dicclair® injections and continued at one week interval for 4 weeks.

This involved a gentle massaging of the abdomen while the opposite hand simultaneously stroked the lower back and tail feathers of a restrained cock. When phallic tumescence was achieved, the collector's hands were placed around the cloaca with a downward pressure while the lower hand exerted slight upward pressure. The semen which pools on the phallus after each squeeze of the cloaca, was collected into a clean dry test tube. To minimize the spread of pathogens, care was taken by the collector not to touch the cloaca structure.

Semen evaluation was done as promptly as possible post collection as described by Rodriguez-Martinez and Barth (2007) for qualitative and quantitative parameters such as semen volume, sperm concentration, sperm motility, semen colour, semen pH, dead sperm percentage and live sperm percentage.

Testicular morphometry

Three cocks in each treatment group were slaughtered and via cervical dislocation and dissection using surgical blade, forceps, scissors, organ weights were measure in grams using a sensitive weighing balance. The testicular organ parts that were measured include; the left and right testis, paired testes, left and right epididymis, paired epididymis, left and right vas deferens, paired vas deferens. At the end, the mean average results of the testicular organ weights from each treatment group was determined.

Hormonal assay

Blood samples (5ml each) were obtained by wing vein puncture of the 20 cocks on day 7 after the Dicclair® injection, for testosterone, FSH and LH evaluation. It was cooled immediately in iced water and transferred to the laboratory, refrigerated at 4°C for 1 hour and the serum separated by centrifugation at 5,000rpm for 10 minutes. The serum was stored immediately at -20°C until enzyme immune assayed (EIA) for testosterone, FSH and LH as described by Micallef *et al.* (1995).

Data analysis

Data collected on semen characteristics, testicular morphometry and hormonal profile of mature Harco cocks were subjected to one - way analysis of Variance (ANOVA) using the technique of steel and Torrie (1980). Significant treatment means were separated using Duncan's New Multiple Rande Test as described by Obi (1990).

RESULTS AND DISCUSSION

The results of Dicclair® administration on semen characteristics of Harco cocks are shown in Table 3.

There were significant differences ($P < 0.05$) among the treatment groups in semen volume, pH, individual motility, sperm concentration, proportions of dead and live sperm cells. Cocks on T₄ recorded the highest value of 1.00ml in semen volume and this differed significantly ($P < 0.05$) from rams on T₂ and T₃ which were similar ($P > 0.05$) to each other in semen volume. Values of semen volume obtained in this study were higher than the range of $0.24 \pm 0.1 - 0.26 \pm 0.3$ ml reported by Abu *et al.* (2006) in cocks and higher than the range $0.25 \pm 0.2 - 0.31 \pm 0.14$ reported by Iheukwumere *et al.* (2008) in Nigerian local cocks. Volume of semen varies with species, age, breed, season and frequency of ejaculation (Ozkan *et al.*, 1998).

Cocks on T₄ recorded the highest value of 8.00 in semen pH and this differed significantly ($P < 0.05$) from cocks on T₁, T₂ and T₃. Cocks on T₁ and T₂ were similar ($P > 0.05$) in PH but differed significantly ($P < 0.05$) from cocks on T₃. The lowest value in pH (6.00) was observed in cocks on T₃. The pH values obtained in this study were

within the normal range of 7-8 reported by Meacham (2002) except cocks T₃ whose pH value (6.00) was lower than the normal range. The measured pH can depend on the length of time since ejaculation and it tends to increase shortly after ejaculation as a result of loss of CO₂ (Meacham, 2002).

Cocks on T₄ recorded the highest value of 82.60% in individual motility and this differed significantly ($P < 0.05$) from cocks on T₁, T₂ and T₃ which were also significantly ($P < 0.05$) different from each other in individual motility. The lowest value in individual motility was observed in cocks on the control treatment (T₁). The values for percentage of motile spermatozoa obtained in this study were higher than the ranges of 36 – 52% and 48.3 – 57.3% reported by Abu *et al.* (2006) and Ameh (2004) respectively in Nigerian cocks. The differences observed in sperm motility may be attributed to breed (Oguike, 2000). Cold shock could not have been responsible for the relatively low motility observed when lower doses of the drug were administered, because estimation of the progressive motility was done soon after semen collection.

Cocks on T₄ recorded the highest value of 1.54 ($\times 10^9$ /ml) in sperm concentration and this differed significantly ($P < 0.05$) from cocks on T₁, T₂ and T₃. Cocks on T₂ and T₃ were similar ($P > 0.05$) to each other, but they differed significantly ($P < 0.05$) from cocks on T₁. The lowest value in sperm concentration was observed in cocks on T₂ and T₃ (0.52×10^9 /ml).

The highest value for sperm concentration obtained in this study (1.54×10^6 /ml) compares favourably with the reports of Abu *et al.* (2006), who recorded 1.98 – 2.10 $\times 10^9$ /ml; Ezekwe *et al.* (2003) who recorded 1.25 – 2.13 $\times 10^9$ /ml and Oguike *et al.* (2000) who reported 1.18 – 2.13 $\times 10^6$ /ml, in Nigerian local cocks but lower than the value of 3.8×10^6 /ml reported by Chalov (1970) in cocks. This variation in sperm concentration of the individual cocks could be attributed to factors such as breed (Oguike *et al.*, 2000), plane of nutrition, ambient temperature, frequency of semen collection and drug administration (Abu *et al.*, 2006).

Cocks on T₁ recorded the highest value of 49.50% in dead sperm cells and this differed significantly ($P < 0.05$) from cocks on T₂, T₃ and T₄ which were similar ($P > 0.05$) to each other in proportion of dead sperm cells. The lowest value in dead sperm cells was observed in cocks on T₄ (35.00%). The percentages of dead sperm cells obtained in this study (35.00 – 49.50%) were higher than the range of 16.54 ± 0.7 – $20.05 \pm 0.05\%$ reported by Iheukwumere *et al.* (2008), the range of 21.0 – 22.5% reported by Abu *et al.* (2006) and the mean value of 14.25% reported by Ameh (2004) in Nigerian local cocks. Neck abnormalities were numerous, followed by tail and mid piece abnormalities. Iheukwumere *et al.* (2008) pointed out that the neck is the most fragile part of the spermatozoa and may be broken while handling semen when making smears.

Cocks on T₄ recorded the highest value of 65.00% in live sperm cells and this differed significantly ($P < 0.05$) from cocks on T₁. There were no significant differences ($P > 0.05$) among cocks on T₄, T₃ and T₂ in proportion of live sperm cells. The lowest percentage of live sperm cells (50.50%) was observed in cocks on the control treatment

(T₁). The highest percentage of live sperm cells obtained in this study (65.00%) was higher than the range of 44.2 – 59.2% reported by Abu *et al.* (2006) and higher than the range of 51.3-4.4% reported by Iheukwumere *et al.* (2008), but lower than the average value of 79.82% reported by Oguike *et al.* (2000) in Nigerian local cocks. It is suggested that high live sperm cells are vital for high fertility (Abu *et al.*, 2006).

Table 1: Doses of Diclair® administered to mature Harco Cocks

Day	Treatment (Diclair® i.u)			
	T ₁	T ₂	T ₃	T ₄
1	0.00	0.03	0.06	0.09
2	0.00	0.03	0.06	0.09
3	0.00	0.03	0.06	0.09
Total	0.00	0.09	0.18	0.27

Table 2: Concentration of Diclair® on mature Harco Cocks

Day	Concentration of Diclair® (i.u)			
	T ₁	T ₂	T ₃	T ₄
1	0.00	4.50	9.00	13.50
2	0.00	4.50	9.00	13.50
3	0.00	4.50	9.00	13.50
Total	0.00	13.50	27.00	40.50

The observation in this study that the group that received higher dose of the test drug recorded the highest percentage of live sperm cells and lowest percentage of dead sperm cells suggests that a higher dose of the drug such as 40.50i.u/cock with 3 days could have high capacity for induction of spermatogenesis and have no deleterious effects on sperm cells. The results of Diclair® administration on testicular morphometry of mature Harco cocks are presented in Table 4.

There were significant differences ($P < 0.05$) among the treatment groups in all the testicular parameters measured.

Cocks on T₄ recorded the highest values in right testis (13.20g) left testis (13.309) and paired testes (25.90g) weights. The lowest values in these parameters were observed in cocks on T₂; 10.15g for right testis, 11.95g for left testis and 22.10g for paired testis weights.

Cocks on T₄ recorded the highest values in right, left and paired epididymal weights, 0.90g, 0.80g and 1.60g respectively. The lowest values in right, left and paired epididymal weights; 0.40g, 0.40g and 0.90g were observed in cocks on T₁, T₂, T₁ and T₂ respectively.

Cocks on T₄ recorded the highest value in right, left and paired vas deferens weights; 0.80g, 0.70g, and 1.40g respectively. The lowest value in right, left and paired deferens weights; 0.50g, 0.50g, and 1.10g were observed in cocks on T₃, T₁, and T₃ respectively.

The observation in this study that cocks on T₄ recorded the highest values in testicular, epididymal and vas deferens weights suggests that 40.50i.u Diclair® treatment could have increased metabolism and efficient utilization of nutrient as well as improved the function of androgen (testosterone) responsible for the development of male reproductive tract (Brackett, 2005).

The results of Diclair® administration on hormonal profile of mature Harco cocks are presented in Table 5. There were significant differences ($P < 0.05$) among the treatment groups in all the hormonal parameters measured; FSH, LH or ICSH, and testosterone.

Table 3: Effect of Diclair® on semen characteristics of mature Harca Cocks

Parameters	Treatment (Diclair® i.u)				SEM
	T ₁ 0.00	T ₂ 13.50	T ₃ 27.00	T ₄ 40.50	
Colour	Creamy-white	Creamy white	Creamy white	Creamy white	Creamy white
Semen volume (ml)	0.88 ^a	0.56 ^b	0.48 ^b	1.00 ^a	0.05
pH	7.00 ^b	7.00 ^b	6.00 ^c	8.00 ^a	0.22
Individual motility (%)	7.00 ^d	77.00 ^b	75.00 ^c	82.60 ^a	0.79
Sperm concentration (x10 ⁹ /ml)	1.14 ^b	0.52 ^c	0.52 ^c	1.54 ^a	0.06
Proportion of dead sperm cells (%)	49.50 ^a	39.60 ^b	38.00 ^b	35.00 ^b	3.14
Proportion of live sperm cells(%)	50.50 ^b	60.40 ^a	62.00 ^a	65.00 ^a	3.14

^{abcd}: Means within row having different superscript are significantly (P<0.05) different. SEM = Standard error of means.

Table4: Effect of Diclair on Testicular Morphometry of Mature Harco Cocks

Parameters	Treatment (Diclair® i.u)				SEM
	T ₁ 0.00	T ₂ 13.50	T ₃ 27.00	T ₄ 40.50	
Right testis weight (g)	11.00 ^b	10.30 ^b	10.15 ^b	13.20 ^a	0.70
Left testis weight (g)	12.70 ^{ab}	12.40 ^b	11.95 ^b	13.30 ^a	0.28
Paired testis weight (g)	24.30 ^a	22.70 ^b	22.10 ^b	25.90 ^a	0.85
Right epididymis weight (g)	0.40 ^c	0.50 ^{bc}	0.80 ^{ab}	0.90 ^a	0.12
Left epididymis weight (g)	0.50 ^{bc}	0.40 ^c	0.70 ^{ab}	0.80 ^a	0.09
Paired epididymis weight (g)	0.90 ^b	0.90 ^b	1.60 ^a	1.60 ^a	0.20
Right vas deferens weight (g)	0.70 ^a	0.70 ^a	0.50 ^b	0.80 ^a	0.06
Left vas deferens weight (g)	0.50 ^b	0.60 ^{ab}	0.60 ^{ab}	0.70 ^a	0.04
Paired vas deferens weight (g)	1.30 ^a	1.30 ^a	1.10 ^b	1.40 ^a	0.06

^{abcd}: Means within row having different superscript are significantly (P<0.05) different. SEM = Standard error of means.

Table 5: Effect of Diclair® on hormonal profile of mature Harco Cocks

Parameters	Treatment (Diclair® i.u)				SEM
	T ₁	T ₂	T ₃	T ₄	
	0.00	13.50	27.00	40.50	
FSH (iu/L)	3.30 ^c	4.18 ^c	25.90 ^a	6.36 ^b	0.31
LH (iu/L)	4.26 ^c	5.16 ^c	37.50 ^a	14.46 ^b	0.71
Testosterone (mg/ml)	2.86 ^c	5.74 ^b	12.66 ^a	1.98 ^c	0.20

^{ab}: Means within row having different superscript are significantly (P<0.05) different. SEM = Standard error of means.

Cocks on T₃ recorded the highest value of 25.90(i.u/L) in FSH and this differed significantly (P<0.05) from cocks on T₁, T₂ and T₄. Cocks on T₁ and T₂ were similar (P>0.05) to each other, but they differed significantly (P<0.05) from cocks on T₄ in FSH values. The FSH values obtained in this study were higher than the range of 0.08 ± 0.08i.u/L – 1.34 ± 0.01i.u/L reported by Iheukwumere *et al.* (2008) in Nigerian local cocks. This may be attributed to differences in breed and drug administration (Culbner *et al.*, 1977; Herbert *et al.*, 2002). Follicle stimulating hormone has been associated with the control of seminiferous tubule growth and differentiation (Culbner *et al.*, 1977; Iheukwumere *et al.*, 2008).

Cocks on T₃ recorded the highest value of 37.50(iu/L) in luteinizing hormone (LH) and this differed significantly (P<0.05) from cocks on T₁, T₂ and T₄. Cocks on T₁ and T₂ were similar (P>0.05) to each other, but they differed significantly (P<0.05) from cocks on T₄ in LH values.

The LH values obtained in this study were higher than the range of 0.65 ± 0.04 i.u/L – 1.36 ± 0.04i.u/L reported by Iheukwumere *et al.* (2008) in Nigerian local cocks. This may be attributed to differences in breed and drug administration (Culbner *et al.*, 1977; Herbert *et al.*, 2002). Luteinizing hormone (LH) or ICSH stimulates the interstitial cells of leydig to produce testosterone which facilitates the process of spermatogenesis (Herbert *et al.*, 2002).

Cocks on T₃ recorded the highest value of 12.66ng/ml in testosterone and this differed significantly (P<0.05)

from cocks on T₁, T₂ and T₄. Cocks on T₁ and T₄ were similar (P>0.05) to each other, but they differed significantly (P<0.05). from cocks on T₂ in testosterone values. The testosterone values obtained in this study were higher than the range of 0.15 ± 0.01ng/ml – 0.21 ± 0.21ng/ml reported by Iheukwumere *et al.* (2008) in Nigerian local cocks. This may be attributed to differences in breed and drug administration (Culbner *et al.* 1977; Herbert *et al.*, 2002).

Conclusion

From the results of the study, it can be concluded that Diclair® improved semen characteristics and testicular, epididymal and vas deferens development of Harco cocks at the level of 40.50i.u, without any deleterious effects on hormonal profile of the cocks.

REFERENCES

- Abu AH, M Ameh and FC Iheukwumere, 2006. Semen Quality of Nigerian Local Cock Reared with Human Menopausal Gonadotrophin (Pergonal®). Livestock Research for Rural Development.
- Ameh M, 2004. Effect of Pergonal® on Semen Quality, Haematological values and carcass characteristics of the Nigerian Local Cocks. M.Sc. Thesis Department of Animal Science and Fisheries, Abia State University Umuahia, Nigeria.
- Bracket BG, 2005. Physiology of Domestic Animals Twelfth Edition William, O. Reece Editor, pp: 670-691.
- Chalov, 1970. Semen Quality and Fertilizing capacity of cocks piferodstron, 20: 24-26.
- Culbner JP, T Sharp and JWC Wells, 1977. Concentration of Testosterone and LH in the blood before and after the onset of spermatogenesis in the Cockerel Report. Fertile, 51: 153.
- Dixon TA and GJ Hopkins, 1996. Super Ovulation in cattle using porcine pituitary Gonadotrophin

- preparation (Plusset Serono) in: Plusset Scientific Literature serono veterinary, Rome, Italy, pp: 22-23.
- Ezekwe AG, IJ Udozor and CO Osita, 2003. Effect of Quantitative feed Restriction on semen Qualities of Nig. Local Cocks Nig. J Anim Prod, 30: 127-132.
- Herbert U, P Okoro, DO Umesiobi and MU Iloeje, 2000. Effects of two preparations of clomiphene citrate on the super ovulation of West African Dwarf Ewes. 14th Inter Congr Anim Reprod, Sweden.
- Herbert U, AH Ezeobi and MU Iloeje, 2002. Induction of spermatogenesis in Rabbits using the fertility Drug Clomiphene citrate (clomid[®]), Proc. 27th Ann. Conf. NSAP, Akure, March 17-27.
- Iheukwumere FC, U Herbert and MU Iloeje, 2004. Haematological and serum Biochemical values of West African Dwarf Does following FSH + LH (Pergonal[®]) Treatment. Int J Agric Rural Dev, 5: 54-60.
- Iheukwumere FC, 2005. Super ovulation in goats in: Afam Anene and Nwaigbo LC (eds.) Issues in sustainable Agriculture in Nigeria. Osprey publication centre, Owerri, Nigeria, 1-9.
- Iheukwumere FC, AH Abu and EC Ndubuisi, 2008. Effect of FSH + LHC (Pergonal[®]) treatment on haematology, immune status and serum metabolites of West African Dwarf Goats. J Anim Vet Adv, 7: 46-50.
- Meacham R, 2002. Perspectives and Editorials. Andrologia. JA Androl, 23: 330-331.
- Micallef IA, MM Hays, A Latif, R Alhasan and SB Sufi, 1995. Serum binding of steroid Tracers and its possible Effects on Direct steroid Immuno assay. Anim Clin Biochem, 32: 566-574.
- Obi IU, 1990. Statistical methods of Detecting differences between Treatment means. Snap press 2nd Ed. Enugu. Nigeria, 24-35.
- Oguike MA, AN Ndubueze and SN Ibe, 2000. Semen Quality of Different Genotypes of Nig. Local cocks. J Sustain Agric Environ, 2: 310-313.
- Ozkan S, P Settari and S Yalun, 1998. Effect of seasonal Ambient Temperature on carcass characteristics of Naked Neck and Normal Feathering Bird. Anim Breed Abstract, 66-361.
- Rodriguez-Martinez H and AD Barth, 2007. In vitro Evaluation of sperm Quality Related to in vivo function and fertility in: Reproduction in Domestic Animals VI Edited by JI Huengel. JE Murray and Mi Smith. Nottingham University press. Nottingham UK. pp: 39-54 SOC Reprod Fert, 64: 39-54.
- Steel RGD and JH Torrie, 1980. Principles and procedures of statistics. A. Biometric Approach 2nd Ed. Mc. Graw-Hill Book Co. Inc. New York.