



Comparative Anatomical, Light and Scanning Electron Microscopical Studies Between Intromittent and Non-Intromittent Typed Phallus of Domestic Goose (*Anser anser domestica*) and Turkey (*Meleagris gallopavo domestica*)

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Article History: 20-073 Received: March 22, 2020 Revised: June 04, 2020 Accepted: June 17, 2020

ABSTRACT

The work applied on the phallus of the adult goose and turkey to give knowledge for the phallus functional morphology and the mechanism of copulation of these domestic birds. It helped the surgical interfering of the wild geese and the artificial insemination in the turkey for commercial production. The phallus of the goose and the phallic bodies of the turkey were demonstrated by anatomical, Histological, histochemical and scanning electron microscopy to compare the micromorphological features. The goose has an intromittent type phallus. It consisted of inner glandular part and outer cutaneous one. The former lined by mucous secretory cells, while the later cover externally by stratified squamous non-keratinized epithelium. The turkey phallus was a non-intromittent type. It composed of a median phallic furrow on the crest at the ventral vent lip and flanked on either side by lateral phallic bodies. The later lined by stratified squamous non-keratinized and supported by longitudinal oriented skeletal muscle which circular in the furrow between two phallic bodies. Additionally, lymphatic aggregation observed in phallus of two birds. This study helped in comparative studies and surgical operations.

Key words: Phallus, Copulatory organ, Goose, Turkey, Scan electron microscope

INTRODUCTION

Several studies applied by many authors on the male reproductive system of the fowl (Marvan, 1969; Lake, 1971; Tingari, 1971; Amer & Shahin, 1975 and King, 1986). The avian male external genitalia showed a great anatomical variation in size and shape even between closely related species. All birds reproduced by internal fertilization except the phallus of some species of birds could be capable of intromission, so the phallus had a variety of forms according to its anatomical structures (Hosken and Stockley, 2004).

The avian phallus divided into three categories, the first was the intromittent type that divided into two types according to presence or absence of a blind tubular cavity within the phallus. The blind cavity absents in kiwis and ostrich, while it present in duck and goose, this type of phallus entered the female cloaca during coitus. The second was the non-intromittent type, which was a reduced type but not lost, as in the fowl and turkey and some song birds, this type did not entered the female cloaca but deposited the semen outside on the female external genitalia. The third

one involved a complete absence of the phallus as in pigeon (Briskie and Montgomerie, 1997; Chen, 2005; Brennan, et. al., 2008 and Herrera et al., 2013).

MATERIALS AND METHODS

Animal ethics

All animal related procedures in the study were approved ethically by Cairo University institutional animal care and use committee (CU-IACUC). Approval document number is CU-11-F-32-19. The study applied on seemingly healthy adult ten male native geese and turkeys, weighting 4.5-6 kg, 3.5-5Kg respectively; they obtained from the chicken farmhouses around Cairo, Egypt.

Gross anatomical study

The phallus was seen insitu at the ventral lip of the vent in the turkey, while laid in a phallic pouch close to cloaca in goose. By using 50 cm syringe filled with warm saline solution, its nozzle was introduced into the phallic pouch and injected with 10% normal saline solution, the phallus could be everted outside the pouch, passed into caudal direction,

Cite This Article as: El-Bably SH and S Hussien, 2020. Comparative anatomical, light and scanning electron microscopical studies between intromittent and non-intromittent typed phallus of domestic goose (*Anser anser domestica*) and turkey (*Meleagris gallopavo domestica*). Int J Vet Sci, x(x): xxxx. www.ijvets.com (©2020 IJVS. All rights reserved)

through the vent, the length could be measured during erection by using a ruler, also could be demonstrated the morphology and different parts by Reilly, 2001. Sony digital camera 24 mp, 18x was used for photographed the existing results. The nomenclature recommended in this study was given by *Nomina Anatomica Avium*.

General histological study

After complete anesthesia, the birds were slaughtered and the phallus of both birds immediately dissected out and sectioned into small pieces. The specimens fixed in neutral buffered formalin. They dehydrated in ascending degree of alcohols, cleared in benzene finally embedded in paraplast. Serial and step serial sections of 5-6 μm thick obtained and stained with Haematoxylin and Eosin (H&E), Masson's trichrome (Bancroft and Gamble, 2008).

Histochemical study

The specimens obtained, fixed and processed as listed in general histological study and Periodic Acid Schiff technique (PAS) stained for neutral mucopolysaccharides detection Carson (1990).

Scanning electron microscope

Small specimens taken from phallus of geese and turkeys washed by 0.1 M Na-cacodylate buffer. They fixed in a mixture of 2.5% paraformaldehyde and 2.5% glutaraldehyde in 0.1 M Na-cacodylate buffer, pH 7.3 for 4 hours at 4 °C thereafter, they washed in the same buffer used and post-fixed in 1% osmic acid in 0.1 M Na-cacodylate buffer for further 2 hours at room temperature. The samples dehydrated and critical point dried with a polaron apparatus. Finally, they coated with gold and observed with JEOL scanning electron microscope (JSM-5400 LV) at KV 10 Polák et al., (1983). All specimens were prepared and analyzed in electron microscope Unit, of applied center for entomonematodes, faculty of Agriculture, Cairo University, Egypt.

RESULTS

The phallus was the organ of copulation in the male's birds, could be categorized into two types, true intromittent phallus, which was introduced into the female cloaca during coitus and non-intromittent phallus was not entered the cloaca but it expelled the sperms onto the external female genitalia directly.

Goose anatomical finding

The phallus was an intermittent type (phallus protrudens). It was a spiral-coiled organ. At the rest state, the phallus of goose invested within a double peritoneal membrane called phallic pouch, slightly sited left to median plane close to the cloaca. It was about 2 and half coils, organized in an anticlock wise direction (Fig. 1A/1) and these coils arranged as a double U-shaped tube that formed of a thick, large part twisted by a small, thin tube, in between the phallic flexure (flexura phalli) (Fig. 1B/ 2). The large thick part called cutaneous sac (sacci cutanei phalli) (Fig. 1B/ 3), and the small thin one called glandular sac (sacci glandularis phalli) (Fig. 1B/ 4). At erected state the phallus was a very long structure composed of base, body and apex, its length about 8-10 cm. The shaft of the

phallus (Fig. 1B/ 5), demarcated by a phallic sulcus (sulcus phalli) was a spirally longitudinal groove, placed between two tubes of the phallus, extended from the ejaculatory fossa to the apex of the phallus called (sulcus spermaticus) (Fig. 1B/ 6). Also, the body characterized by presence of phallic ridges spread all over the phallus, arranged in a numerous row, these ridges were pointed and directed backward towards the base called (rugae phalli) (Fig. 2/ 1). The base of the phallus (Fig. 3A) supported by a cartilagenous body (corpus cartilagenous) (Fig. 3B/ 4). The suspensory ligament of the phallus called suspensorium phalli (Fig. 3C/ 6). The apex of the phallus (apex phalli erecti) was the terminal part of the phallus (Fig. 3D/7), it had the opening of glandular sac (ostium sacci glandularis phalli) that allowed the seminal fluid to be come out from the phallus (Fig. 3D/ 8).

Goose histological finding

histologically, the phallus of goose consist of the inner glandular part and outer cutaneous layer. The inner glandular part lined with mucous secretary cells and contained mucous gland (Figs. 4A, 4B and 4C) whose revealed PAS positive reaction (Fig.4D) and surrounded by outer cutaneous layer of collagen fibers (Fig. 5A); the latter layer composed of two distinct parts: an inner layer of loose circumferential and an outer dense disorganized layer in between the two layers was large lymphatic lumen (Fig. 5B), as well as lymphatic capillaries observed in the connective tissue beneath the glandular portion (Fig. 4C). The surface epithelium of its cutaneous part lined by stratified squamous non keratinized epithelium Fig. (5C). Numerous lymph nodules observed under the epithelium (Fig.5D). The base of the phallus was supported by hyaline cartilage, lymphatic capillaries and skeletal muscle (Fig. 5E).

Turkey anatomical finding

The male copulatory organ appeared in the ventral wall of the vent; it was non intromittent type. It was composed of two lateral phallic bodies on each side at the ventral wall of the vent (Fig. 6/1), separated by a median furrow or groove (Fig. 6/2). Two lateral phallic bodies appeared as a short, rounded structure, which was enlarged and engorged with fluid; the fluid passed into the median furrow.

Turkey histological finding

The phallus of turkey formed of cutaneous portion only (Fig. 7A) lined with stratified squamous non keratinized epithelium (Fig. 7B) and characterized by neither tubular nor glandular portion detected. This cutaneous portion arranged as two plugged portions, lateral phallic bodies (Fig. 7A), connected by furrow. Longitudinal oriented skeletal muscle observed in the phallic body while circular orientation in the furrow (Figs. 7C and 8A). Highly vascularized and innervated dense irregular connective tissue mainly collagen fibres with peripheral nerve ending, lamellar corpuscle, observed beneath the stratified squamous non keratinized epithelium (Figs. 7C and 8B). Lymphatic nodules detected beneath the epithelium in the phallic bodies as well as at commissure between the phallic bodies and the furrow (Figs. 8C and 8D).

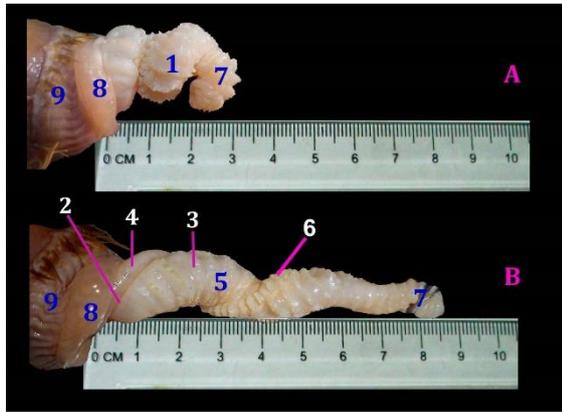


Fig. 1: A photograph showing the length and parts of erected phallus of the male goose. A- Length of the flaccid phallus. B- Length of the fully erected phallus. 1- Anticlock wise coils, 2- Phallic flexure, 3- Cutaneous sac. 4- Glandular sac, 5- Body, 6- Spermatic sulcus, 7- Apex, 8- Base, 9- Vent.

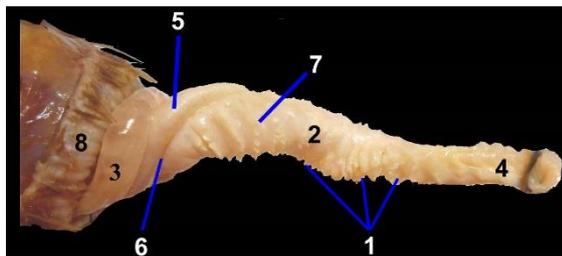


Fig. 2: A photograph showing a fully erected goose phallus. 1- Phallic ridges, 2- Body, 3- Base, 4- Apex, 5- Glandular sac, 6- Phallic flexure, 7- Cutaneous sac, 8- Vent.

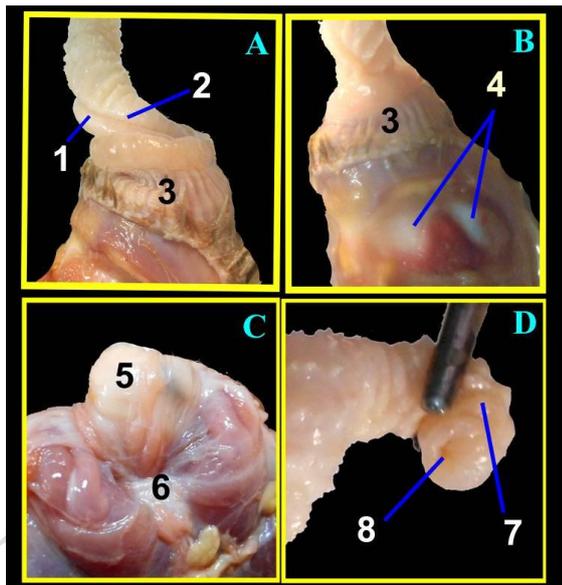


Fig. 3: A photograph showing the different parts of the phallus in the male goose. A- Base, B- cartilaginous body, C- Phallic pouch, D- Apex and opening of glandular sac. 1- Glandular sac, 2- Cutaneous sac, 3- Vent, 4- Hyaline cartilage, 5- Phallic pouch, 6- Suspensory ligament, 7- Apex, 8- Opening of glandular sac

DISCUSSION

The present study revealed that, the phallus of the goose was an intromittent type, it was a spiral coiled structure invested within a double peritoneal membrane (Phallic pouch), similar to Getty (1975), King (1981),

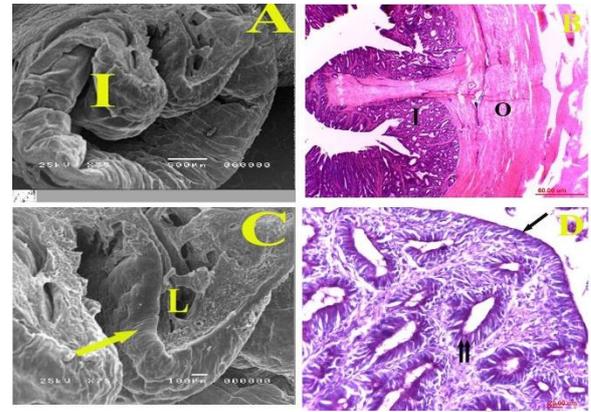


Fig. 4: A transverse section of goose phallus, A- Scanning electron microscopy showing inner glandular part (I). B- A photomicrograph showing inner glandular part (I), outer collagenous part (O). H&E stain (X40). C- Scanning electron microscopy showing mucous secretory cells (arrow) & lymphatic capillaries (L). D- A photomicrograph showing inner glandular part lined with PAS positive mucous secretory cells (arrow) and mucous gland (double arrow). PAS stain (X400)

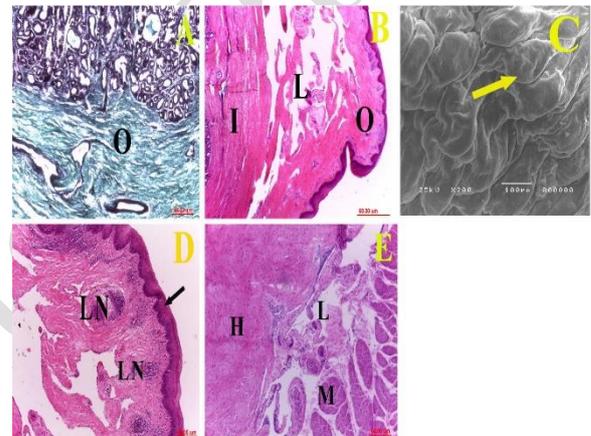


Fig. 5: A transverse section of goose phallus, A- A photomicrograph showing outer cutaneous layer of collagen fibers (O). Masson's trichrome stain (X100). B- A photomicrograph showing an inner layer of loose circumferential connective tissue (I) & an outer dense disorganized one (O) in between the two layers was large lymphatic lumen (L). H&E stain (X40). C- Scanning electron microscopy showing the surface epithelium of its cutaneous part lined by stratified squamous non keratinized epithelium (arrow). D- A photomicrograph showing a numerous lymph nodules (LN) observed under the stratified squamous non keratinized epithelium (arrow). E- A photomicrograph of goose phallus base showing hyaline Cartilage (H), lymphatic capillaries (L) and skeletal muscle (M). H&E stain (X100)

Mc Lelland (1990), Kevin (2000), Brennan et al. (2009) and El Gindy et al. (2016) in duck and King and Mc Lelland (1984), Baumel et al. (1993), Brennan and Prum (2012) in Ostrish and kiwis and Rajendranath et al. (2013) in emu, while, the phallus of the turkey was an non intromittent type and lied on the crest of the ventral lip of the vent, these results similar to the results in the domestic fowl that confirmed by Bull et al. (2007).

The coils of phallus were about 2 and half coils, had a anticlock wise direction, but El Gindy et al. (2016) in balady duck recorded 3 and half coils. When dissected it was seen as a U-shaped tube, attached to the cloaca at both

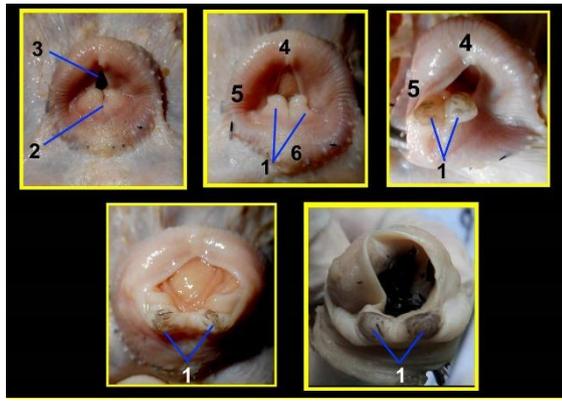


Fig. 6: A photograph showing the phallus in the male Turkey. 1- Lateral phallic bodies, 2- Median furrow, 3- Opening of the vertical fissure, 4- Dorsal lip, 5- Vent, 6- Ventral lip.

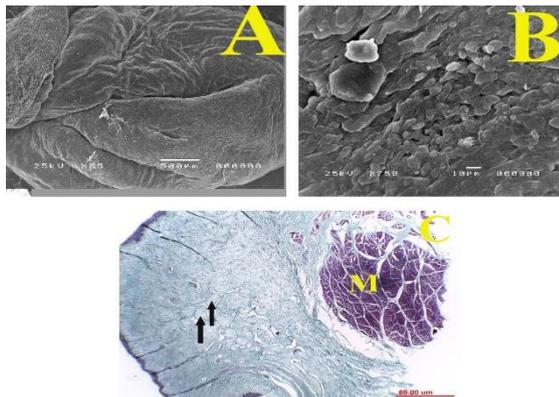


Fig. 7: A transverse section of turkey phallus. A- Scanning electron microscopy showing the cutaneous portion in the phallic body. B- Scanning electron microscopy showing the cutaneous portion lined by stratified squamous non keratinized. C- A photomicrograph showing longitudinal oriented skeletal muscle (M) & lamellar corpuscle (arrow) in the phallic body. Masson's trichrome stain (X40).

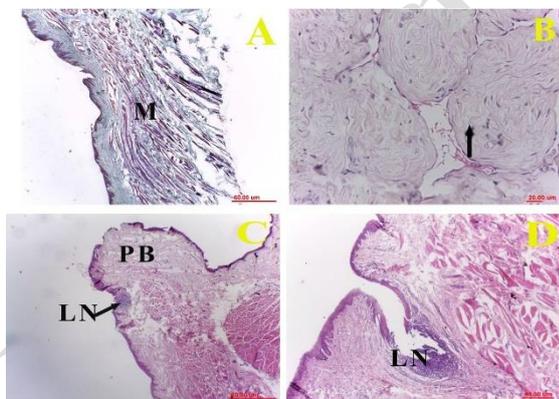


Fig. 8: A transverse section of turkey phallus. A- A photomicrograph showing circular oriented skeletal muscle (M) in the furrow. Masson's trichrome stain (X40). B- A photomicrograph showing lamellar corpuscle (arrow) in the phallic body. H&E stain (X400). C- A photomicrograph showing lymphatic nodule (LN) in the phallic body (PB). H&E stain (X40). D- A photomicrograph showing Lymphatic nodule at the commissure between the phallic body and the furrow (LN). H&E stain (X100).

ends that simulated Kevin (2000) in Argentine duck and El Gindy et al. (2016) in balady duck and. This tube formed of two parts; thick and thin tubes in between phallic flexure,

the thick part called cutaneous sac while, the thin one called glandular sac similar to King and Mc Lelland (1984), Baumel et al. (1993) and El Gindy et al. (2016) in duck, but in this study, the two tubes in the goose were a very characteristic than the same tubes in the duck as the cutaneous tube was a very larger than the glandular one.

In the turkey, the phallus formed of two lateral phallic bodies on each side at the ventral wall of the vent. Two lateral phallic bodies appeared as a short, rounded structure, separated by a median furrow, these results confirmed by Bull et al. (2007) in fowl but the author reported presence of a median phallic body between the two phallic bodies, which was not found in the turkey in this study and also stated that, the deposited semen became outside the female cloaca, on the everted vagina during copulation process, then the spermatozoa transported into the oviduct. In goose, the large cutaneous sac of phallus characterized by a spirally longitudinal phallic sulcus that extended from the base to the apex, also called sulcus spermaticus due to the sperms transported through this groove called sulcus spermaticus that similar to King (1981), Baumel et al., (1993), Duan (2000), El Gindy et al. (2016) and Hassan (2016) in balady duck, Wang et al. (2008) in goose, while Yong and Zhanjun (2011) in ostrich recorded a penile groove from at the back of the penis which was a closed and like a tube when the penis erected.

In this study, the fully erected phallus of goose consisted of a large left and a small right tubes twisted around each other, which formed a bulbous base, body and terminated by the apex of the phallus, as recorded by Getty (1975), King and Mc Lelland (1984) and Mc Lelland (1990) in duck these regions founded in the phallus of all types of Anseriforms but the divisions of these regions differed according to each author, as Brennan et al. (2009) in Muscovy duck divided it into basal and apical regions, Baumel et al. (1993) in duck demonstrated four components, phallic base, phallic body, phallic sac and phallic pouch. While in the turkey, the phallus composed of a median phallic groove flanked on either side by lateral phallic bodies. Regarding the total length of the phallus, was ranged from 8-10 cm during fully erection in goose, while, reached 22 cm in Argentine duck (Kevin, 2000), 20 cm in ostrich (Elias et al., 2007 and Brennan and Prum, 2012), 25-30 cm in ostrich (Yong and Zhanjun, 2011) who stated that this penis was a very strong, while the penis of the turkey as in chickens was not strong, 3 cm. in rhea (Góes et al., 2010), 8.5 cm in Laysan ducks and 5.3 cm in Mandarin ducks (Herrera et Al., 2014) and ranged from 13-15 cm in balady duck (El Gindy et al., 2016 and Hassan, 2016), The phallus had some black coloration on its outer surface in Mandarin ducks and also lacked this coloration in the Laysan ducks that's recorded by (Herrera et Al., 2014), this coloration not recorded in the goose in this study and also not recorded by El Gindy et al. (2016) and Hassan (2016) in balady duck. The external surface of the phallus characterized by presence of ridges that covering the wall of the phallus, as Brennan et al., (2009) in mallard duck, El Gindy et al. (2016) and Hassan (2016) in balady duck but this study added that these phallic ridges were pointed, distributed all over the phallus in a several rows. Meanwhile Kevin (2000) in Argentine duck reported spines directed backwards towards the base of the phallus.

The penis of goose had the inner glandular part lined with secretory cells surrounded by outer cutaneous layer of collagen fibers; the latter layer composed of two distinct parts: an inner layer of loose circumferential and an outer dense disorganized layer in between the two layers was large lymphatic lumen. This result agreed with Brennan et al., (2009) in mallard duck, Brennan and Prum (2012) in emus, Hassan (2016) and El Gindy et al. (2016) in balady duck, while Brennan and Prum (2012) in Ostrich recorded that, the penis lacked a blind tubular cavity and formed of fibrous bodies composed of dense, largely disorganized collagen matrix except in the areas surrounded the narrow lymphatic spaces.

In line with King and Mc Lelland (1984), Baumel et al. (1993), Brennan et al. (2009) in duck and El Gindy et al., (2016) in balady duck, the base of phallus was supported by cartilaginous body which formed of hyaline cartilage and its cutaneous part.

In the present study lymphocytic aggregations noticed in the phallus of goose and turkey. Lymphocytic aggregation had developed a number of different immunological strategies including cell mediated one. These lymphocytes might add a more protective condition for the sperms. In the current study, PAS reaction detected in the surface epithelium and the gland. This revealed presence of neutral mucopolysaccharides Zaher et al. (2012).

Conclusions

The study gives a complete knowledge for phallus of geese and turkey that helped in the surgical operations of the wild geese, and the artificial insemination in the turkey, which was an obligatory method for commercial production.

Author contributions

All authors contributed to the reagents/materials/analysis tools, collected the material, analyzed the data and wrote the manuscript.

REFERENCES

- Amer FI and Shahin MA, 1975. The post-hatching development of the gonads in the fowl, *Gallus domesticus*. *Ann. Zool*, 11: 1-25.
- Bancroft JD and Gamble M, 2008. *Theory and practice of histological techniques*. 6th ed. Churchill Livingstone Edinburgh, London and New York.
- Baumel JJ, King SA, Breasile JE, *et al.*, 1993. *Nomina Anatomica Avium*. Published by the Nuttall Ornithological Club. No: 23, Cambridge, Massachusetts.
- Brennan PLR, Birkhead TR, Zyskowski K, *et al.*, 2008. Independent evolutionary reductions of the phallus in basalbirds. *J Avian Biol*, 39: 487-492.
- Brennan PLR, Clark CJ and Prum RO, 2009. Explosive eversion and functional morphology of the duck penis supports sexual conflict in waterfowl genitalia. *Proc R Soc*, 277: 1309-1314.
- Brennan PLR and Prum RO, 2012. The erection mechanism of ratite penis. *J Zool*, 286: 140-144.
- Briskie JV and Montgomerie R, 1997. Sexual selection and the intromittent organ of birds. *J Av Biol*, 28: 73-86.
- Bull ML, MRF Martins, MD Cesario, CR Padovani and AA Mendes 2007. Anatomical study on domestic fowl (*Gallus domesticus*) Reproductive system. *Int Morphol*, 25: 709-716.
- Carson FL, 1990. *Histotechnology: A Self-Instructional Text*. American Society of Clinical Pathologists. Chicago, pp: 294.
- Chen YT, 2005. Matching methods and the relevant management skills of pigeon breeders. *China Poult*, 27: 26-28.
- Duan X, 2000. The main points of artificial insemination. *China Poult*, 22: 17-18.
- EL Gindy EM, Hassan AM, El-Bably SH *et al.*, 2016. Morphological Studies on the Male Copulatory Organ of Balady Duck (*Anas boschas domestius*) with Special Reference to Its blood Supply. *Giza Vet Med J*.
- Elias MZJ, Aire TA and Soley JT, 2007. Macroscopic Features of the Arterial Supply to the Reproductive System of the Male Ostrich (*Struthio camelus*). *Anat Histol Embryol*, 36: 255-262.
- Getty R, 1975. *The anatomy of the domestic animals*. Volume 2. Fifth Edition. W.B. Saunders Company, Philadelphia London.
- Góes PAA, Cavalcante AKS, Nichi M, *et al.*, 2010. Reproductive characteristics of captive greater rhea (*Rhea americana*) males reared in the state of Sao Paulo. *Brazil. Braz J Poult Sci*, 12: 57-62.
- Hassan AM, 2016. *Some Anatomical Studies on the Urogenital System of Balady Duck*. PhD Vet Degree. Faculty of Veterinary Medicine, Cairo University, Egypt.
- Herrera AM, Brennan PLR and Cohn MJ, 2014. Development of Avian External Genitalia: Interspecific Differences and Sexual Differentiation of the Male and Female Phallus. *Sex Dev*, 9: 43-52.
- Herrera AM, Shuster SG, Peritton CL *et al.*, 2013. Developmental Basis of Phallus Reduction during Bird Evolution. *Current Biology*, 23: 1065-1075.
- Hosken DJ and Stockley P, 2004. Sexual selection and genital evolution. *Trends Ecol Evol* 19: 87-93.
- Kevin GMC, 2000. The 20-cm Spiny Penis of the Argentine Lake Duck (*Oxyura vittata*). *The Auk*, 117: 820-825.
- King AS, 1981. Phallus. In form and function in birds. New York. Academic press: pp: 107-148.
- King AS and Mc Lelland J, 1984. *Birds, Their Structure and Function*. 2nd ed, Bailliere Tindall, England: pp: 145-165.
- King AS, 1986. Aparelho urogenital das aves. In: Getty, R. Sisson and Grossman's. *Anatomia dos animais domésticos*. 5th ed. Rio de Janeiro, Interamericana, 2: 1805-13.
- Lake PE, 1971. The male in reproduction. In: Bell, DJ Freeman, BM Physiology and Biochemistry of the Fowl. New York, Academic Press, 3: 1411-47.
- Marvan F, 1969. Postnatal development of the male genital tract of the *Gallus domesticus*. *Anat Anz Bd*, 124:443-62.
- Mc Lelland J, 1990. *A colour atlas of avian anatomy*. Wolfe Publishing Ltd., London, pp: 68-83.
- Polák Š, Brozman M and Jakubovský J, 1983. Concerning problem of applying raster electron microscopy in studying the ortho- and pathomorphology of the human spleen. *Bratislava Lekarske Listy* 80: 267-278.
- Rajendranath N, S Chandrasekhara RT, Kumar PD, *et al.*, 2013. Gross Anatomical Studies on the Apparatus of the Emu (*Dromaius novaehollandiae*). *Indian J Vet Anat* 25: 81-82.
- Reilly JS, 2001. Euthanasia of animals used for scientific purposes; section 13: *Birds (Class Aves)*, ANZCCART, 2nd ed. pp: 79-82.
- Tingari MD, 1971. On the structure of the epididymal region and ductus deferens of the domestic fowl (*Gallus domesticus*). *J Anat*, 109: 423-35.
- Yong Z and Zhanjun R, 2011. Anatomic Study on the Main Male Reproductive Organs of Ostrich, *Global. J Health Sci*, 3: 2011.
- Wang, Wenqiang, Huaqi Z, *et al.*, 2008. Artificial Insemination Technology and the Application in Goose. *China Poult*, 30: 37-38.
- Zaher M, EL_Ghareeb AW, Hamdi H *et al.*, 2012. Histochemical adaptation of avian alimentary canal to their food habits: *I Coturnixcoturnix*. *Life Sci J*, 9: 253-275.