Hepatoprotective Effect of Oyster Mushroom \((Pleurotus Sajor Caju)\) in Broilers Fed Aflatoxin

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**INTRODUCTION**

Aflatoxins are secondary metabolites produced by certain fungi belonging to the genus \(Aspergillus\). Aflatoxins are produced in feed stuff, when appropriate conditions of humidity and temperature exist during storage (Fernandez et al., 1994). Consumption of such toxin contaminated feed poses great hazards to poultry health by causing liver damage, retarded growth, impaired feed conversion and immune suppression through depression of both cell mediated and humoral immunity (Shivachandra et al., 2003).

Six hundred and fifty one species representing one hundred and eighty two genera of hetero and homobasidiomycetes mushrooms have been found to contain antitumor or immune stimulating polysaccharides. There are many other beneficial effects found in mushrooms, such as antioxidant (Lakshmi et al., 2005), hypoglycemic, anti-inflammatory and hepatoprotective activities (Leifa Fan et al., 2000 and Jayakumar et al., 2006).

Hence in the present study, the hepatoprotective effect of \(Pleurotus sajor caju\) was assessed against experimental aflatoxicosis in broilers by evaluating serum biochemical parameters, antioxidant profile and pathological alterations of liver.

**MATERIALS AND METHODS**

The experiment was conducted as per the guidelines of the Institutional Animal ethical committee.

Aflatoxin \(B_1\) was produced on sterile rice using \(Aspergillus parasiticus\) NRRL 2999 strain (Shotwell et al., 1966) and estimated by Romer method (1975) using Thin Layer Chromatography. Oyster mushroom was purchased from a commercial production unit at Coimbatore, India, shade dried and powdered. The toxin and mushroom powder were mixed with feed in appropriate quantities according to experimental design.

Sixty commercial day old straight run broiler chicks (Vencob strain) were weighed, wing banded and reared under standard management conditions. The chicks were fed standard broiler feed free of aflatoxin and water \(ad\ l\ Fibonacci\) during the first week. On day eighth, the chicks were randomly divided into six treatment groups of ten each and the following treatment was given from the eighth day to forty second day.

Group I was fed with normal feed (control). Aflatoxin (1ppm) mixed feed was given to group II (Aflatoxin control). Group III was fed with mushroom (5%) mixed feed (mushroom control). Aflatoxin 1ppm along with 1%, 2.5% and 5% mushroom was mixed with feed and fed to groups IV, V and VI respectively.
The birds were sacrificed on 42nd day of age. Gross pathology of liver was studied. Blood samples were collected for estimation of serum protein, Alanine aminotransferase, Aspartate aminotransferase by colorimetric method (Reitman and Frankel, 1957) and lipid peroxidation by Thiobarbituric acid- reacting substances method (Ohkawa et al., 1979). Liver samples were collected, washed in saline, kept in freezer for the estimation of Glutathione peroxidase (Rotruck et al., 1973) and Superoxide dismutase (Marklund and Marklund, 1974) and in 10% buffered neutral formalin for histopathological studies.

Data were subjected to statistical analysis as per Snedecor and Cochran (1989) by Completely Randomized Design.

RESULTS AND DISCUSSION

Serum biochemical changes

Aflatoxin treatment resulted in highly significant (P ≤ 0.01) decrease in serum proteins, albumin and globulin (Table 1). This decrease might be due to binding of aflatoxin metabolite to DNA, disrupting transcriptional events leading to inhibition of protein synthesis. (Huff et al., 1986)

Mushroom treatment resulted in significant (P ≤ 0.01) ** increase in serum total protein compared to that of group II. The restorative effect exhibited by mushroom may be due to its high protein content (Cohen et al., 2002 and Rajini Goyal et al., 2006). In addition the mushroom also contains basic antioxidant compounds namely ascorbic acid, Vitamin C, Vitamin E, β carotene and phenolic compounds (Yang et al., 2002). Hence the restoration may be due to the hepatoprotective effect of mushroom against aflatoxin B1.

The ALT and AST values were found to be significantly (P≤0.05) elevated in aflatoxin alone fed group (Table 1) indicating interruption in liver function. Mushroom treatment resulted in a dose dependant significant decrease in ALT and AST compared to that of group II. The ability of the mushroom to inhibit CYP1A activities might prevent the epoxide formation from aflatoxin B1, reducing the hepatic damage (Khlood et al., 2005).

Table 1: Effect of oyster mushroom (Pleurotus sajor caju) on serum biochemical parameters and antioxidant profile (mean ± S.E ) in aflatoxin fed broilers (n=6)

<table>
<thead>
<tr>
<th>Group</th>
<th>Total protein** (g/dl)</th>
<th>Albumin** (g/dl)</th>
<th>Globulin* (g/dl)</th>
<th>ALT (SGPT)* Units/ml</th>
<th>AST (SGOT)* Units/ml</th>
<th>Lipid peroxidation* nmol/ml</th>
<th>Glutathione peroxidase** mg/min/mg of GSH consumed</th>
<th>Superoxide dismutase* Units/min/mg of protein</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>5.88±0.13</td>
<td>1.71±0.06</td>
<td>4.17±0.16</td>
<td>13.33±1.23</td>
<td>126.33±5.74</td>
<td>31.20±0.95</td>
<td>18.93±0.09</td>
<td>1.22±0.01</td>
</tr>
<tr>
<td>AFB1 1 ppm</td>
<td>4.83±0.21</td>
<td>1.21±0.04</td>
<td>3.62±0.22</td>
<td>20.00±0.89</td>
<td>148.00±4.47</td>
<td>41.99±4.67</td>
<td>13.76±0.17</td>
<td>0.76±0.01</td>
</tr>
<tr>
<td>Mushroom 5%</td>
<td>5.84±0.16</td>
<td>1.77±0.06</td>
<td>4.07±0.18</td>
<td>14.00±1.46</td>
<td>132.00±6.39</td>
<td>31.63±1.24</td>
<td>18.49±0.17</td>
<td>1.23±0.01</td>
</tr>
<tr>
<td>AFB1 1 ppm +</td>
<td>5.65±0.13</td>
<td>1.53±0.02</td>
<td>4.12±0.13</td>
<td>16.67±1.69</td>
<td>141.67±5.90</td>
<td>33.37±1.24</td>
<td>15.39±0.21</td>
<td>1.03±0.01</td>
</tr>
<tr>
<td>Mushroom 1%</td>
<td></td>
<td></td>
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</tr>
<tr>
<td>AFB1 1 ppm +</td>
<td>5.93±0.13</td>
<td>1.62±0.04</td>
<td>4.31±0.14</td>
<td>15.33±1.33</td>
<td>131.00±6.17</td>
<td>31.63±1.83</td>
<td>17.18±0.21</td>
<td>1.14±0.02</td>
</tr>
<tr>
<td>Mushroom 2.5%</td>
<td></td>
<td></td>
<td></td>
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<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>AFB1 1 ppm +</td>
<td>5.98±0.11</td>
<td>1.63±0.04</td>
<td>4.35±0.13</td>
<td>14.67±1.23</td>
<td>129.00±4.02</td>
<td>31.20±1.50</td>
<td>18.07±0.18</td>
<td>1.20±0.01</td>
</tr>
<tr>
<td>Mushroom 5%</td>
<td></td>
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</tbody>
</table>

*Overall mean bearing different superscripts between rows differ significantly (P ≤ 0.05); **Overall mean bearing different superscripts between rows differ significantly (P ≤ 0.01)

Antioxidant profile

A significant increase in lipid peroxidation (P≤0.05) and decrease in Glutathione peroxidise (P≤0.01) and Superoxide dismutase (P≤0.05) was recorded in aflatoxin alone fed group (Table 1). Among the treatment groups there was a dose dependant significant decrease in lipid peroxidation and increase in SOD and GPx activities. Pleurotus species play a major role in inhibiting lipid peroxidation due to aflatoxin feeding (Nayana Jose and Janardhanan, 2000). In addition to this Pleurotus species contains basic antioxidant compounds namely ascorbic acid, Vitamin C, Vitamin E, β Carotene and phenolic compounds (Yang et al., 2002) which might be responsible for reducing the oxidation process due to aflatoxin feeding.

Gross Pathology

In aflatoxin control group the liver was enlarged, friable and pale. In addition, ascites and enlargement of kidney were observed.

Mushroom treatment at 1% level resulted in moderate enlargement and congestion of liver. Mushroom treatment at 2.5% and 5% resulted in almost normal appearance of liver indicating hepatoprotective effect.

Histopathology

The liver from mushroom control group showed normal histology (Plate I).

The results of histopathological studies of liver from aflatoxin fed group revealed macrovesicular changes, replacement of necrosed hepatocytes by infiltrating mononuclear cells, bile duct hyperplasia (Plate II) and central venous congestion. In addition to these changes, pseudo acini formation and microcyst formation were also noticed.

Among the treatment groups, liver from group IV, showed diffuse necrosis of hepatocytes, pseudo acini formation, infiltration of mononuclear cells, macro and micro vesicular changes, venous congestion and disruption of hepatic cords (Plate III).
While group V showed infiltration of inflammatory cells, focal necrosis of hepatocytes and cloudy swelling of surrounding cells (Plate IV), group VI showed only microvesicular changes and slight disruption of hepatic cords (Plate V).

Among the treatment groups, a dose dependant improvement in the hepatic architecture was observed. *Pleurotus ostreatus* mushroom administration to the rats with hepatic damage induced by CCl₄, resulted in only minimal disruption of hepatic cellular structure (Jayakumar et al., 2006).

These results indicate the hepatoprotective effect of the mushroom. Hence the present study is suggestive of substantial reversal of aflatoxin induced hepatotoxicity by *Pleurotus sajor caju* mushroom in a dose dependant manner.

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